

Safety Data Sheet

Cyclophosphamide

Division of Safety
National Institutes
of Health



WARNING!

THIS COMPOUND IS ACUTELY TOXIC, CARCINOGENIC, TERATOGENIC, EMBRYOTOXIC, AND MUTAGENIC. IT IS READILY ABSORBED BY VARIOUS BODY TISSUES, THROUGH THE SKIN, INTESTINAL TRACT, AND TRANSPLACENTALLY. AVOID FORMATION AND BREATHING OF AEROSOLS.

LABORATORY OPERATIONS SHOULD BE CONDUCTED IN A FUME HOOD, GLOVE BOX, OR VENTILATED CABINET.

AVOID SKIN CONTACT: IF EXPOSED, WASH WITH SOAP AND COLD WATER. AVOID WASHING WITH SOLVENTS. AVOID RUBBING OF SKIN OR INCREASING ITS TEMPERATURE.

FOR EYE EXPOSURE, IRRIGATE IMMEDIATELY WITH LARGE AMOUNTS OF WATER. FOR INGESTION, INDUCE VOMITING. DRINK MILK OR WATER. REFER FOR GASTRIC LAVAGE. FOR INHALATION, REMOVE VICTIM PROMPTLY TO CLEAN AIR. ADMINISTER RESCUE BREATHING IF NECESSARY. REFER TO PHYSICIAN.

IN CASE OF LABORATORY SPILL, WEAR PROTECTIVE CLOTHING DURING CLEANUP. AVOID SKIN CONTACT OR BREATHING OF AEROSOLS. SEE CASTEGNARO ET AL. (1985) FOR DETAILS. DISPOSE OF WASTE SOLUTIONS AND MATERIALS APPROPRIATELY.

A. Background

Cyclophosphamide (CP) in its hydrated (usual) form is a white odorless powder with a slightly bitter taste. It is toxic, carcinogenic, mutagenic, and teratogenic in animals and probably in man. Its mode of action is that of an alkylating agent after activation. Its principal uses are as an antineoplastic agent, as an immunosuppressive agent in a variety of non-malignant diseases, as an insect chemosterilant, and as a defleecing agent for sheep.

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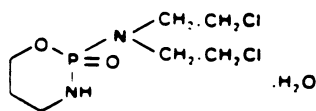
CP has been identified by an IARC Working Group as being probably carcinogenic for humans (Group 2A) (Althouse et al., 1980).

Recent review articles include IARC, 1981; Colvin and Hilton, 1981.

B. Chemical and Physical Data

Introductory notes: (a) The structure of CP reveals an asymmetric phosphorus atom and, therefore, this compound exists as two optical isomers. The levo- and dextro-rotary enantiomers have been identified by x-ray analysis as the S- and R-forms, respectively (Sato et al., 1983). Commercial preparations are unresolved but there are indications that differences exist in the relative abundances of these enantiomers in such preparations (Mruzek and Shaw, 1984). With the exception of a few studies dealing with their optical or biological properties, which are mentioned in the text, no attention has been paid by investigators to the existence of isomers or the isomeric composition of their material. (b) Unless otherwise noted, all data are for the monohydrate which is the usual form.

1. Chemical Abstract Nos.: a. general: 50-18-0; b. levo-CP: 60007-96-7; c. dextro-CP: 60030-72-0; d. racemic CP: 6007-95-6.
2. Synonyms: 2H-1,3,2-oxazaphosphorine, 2-[bis-(2-chloroethyl)-amino]tetrahydro-2-oxide;^A 2H-1,3,2-oxazaphosphorine-2-amine,N,N-bis-(2-chloroethyl)tetrahydro-2-oxide;^B 1-bis(2-chloroethyl)amino- 1-oxo-2-aza-5-oxaphosphoridine monohydrate; [bis(chloro-2-ethyl)-amino]-2-tetrahydro-3,4,5,6-oxazaphosphorine-1,3,2-oxide-2-monohydrate;^C B518; Cyclophosphan(e); Cytosan; Endosan; Genosan; Mitosan; Procytox; Sendosan; NSC 26271.
3. Chemical structure and molecular weight: $C_7H_{15}Cl_2N_2O_2P \cdot H_2O$; M.W. 279.1



4. Density: No data.
5. Absorption spectroscopy: Ultraviolet absorption maximum (log ϵ) at 198 nm (2.667) in methanol (Wantland and Hersh, 1979). The mass spectrum has been reported (Schulten, 1974).

^AChemical Abstracts name, used for listings in 7th and 8th Decennial Index.

^BChemical Abstracts name, used for listings in 9th Decennial Index and subsequently.

^CFor other chemical names see IARC (1981).

Optical rotation: $[\alpha]_D^{25} = +2.48$ and -2.46 for the two optical isomers (Sato et al., 1983).

Volatility: No data; may be assumed to be low.

Solubility: In water and physiological saline, 40 g/L. Soluble in ethanol, methanol, chloroform, dioxane, ethylene glycol; slightly soluble in ether and acetone.

Description: The monohydrate is a white colorless crystalline powder with a slightly bitter taste. The anhydrous material is an oily semisolid.

Boiling point: No data; melting point: variously reported as 1-45°C and 49.5-53°C. (The latter range appears to be more reliable.) Melting point of both optical isomers: 67-68°C.

Stability: Decomposition in aqueous solution (physiological saline) at room temperature for 8 and 24 hours and 1 week is: less than 1.5%, 3.5%, and 11.9%, respectively; less than 1.5% decomposition in the refrigerator in 6 days (Gallelli, 1967; Brooke et al., 1973). Stable (less than 10% change in 24 hours at room temperature) in 5% dextrose in plastic containers (Benvenuto et al., 1981). Hydrolyzes at temperatures above 30°C (Arnold and Klose, 1960; Zon et al., 1971; Kensler et al., 1979). Anhydrous CP decomposes slowly on standing at room temperature. At temperatures above 30°C and relative humidity of less than 70% the hydrate slowly changes to the anhydrous form (Laine et al., 1983).

Chemical reactivity: No data other than those for hydrolysis. Note that liberation of chloride ion in aqueous solution is very much slower than from alkyl nitrogen mustards under comparable conditions. 2-Chloroethylamino compounds are usually subject to reaction with oxidizing agents.

Flashpoint: No data.

Autoignition temperature: No data.

Explosive limits in air: No data.

Fire, Explosion, and Reactivity Hazard Data

CP does not require special fire-fighting procedures or equipment and does not present unusual fire and explosion hazards.

No conditions contributing to instability are known to exist, other than its instability to light, to water at elevated temperatures, and probably to oxidizing agents.

3. No incompatibilities are known.
4. No hazardous decomposition products have been identified. The metabolism of CP results in the production of acrolein, which is a flammable irritant to eyes and mucosa; however, there are no data indicating that acrolein is produced under conditions of fire from the solid material.
5. CP does not require non-spark equipment.

Operational Procedures

The NIH Guidelines for the Laboratory Use of Chemical Carcinogens describe operational practices to be followed when potentially carcinogenic chemicals are used in NIH laboratories. The NIH Guidelines should be consulted to identify the proper use conditions required and specific controls to be implemented during normal and complex operations or manipulations involving CP.

It should be emphasized that this data sheet and the NIH Guidelines are intended as starting points for the implementation of good laboratory practices when using this compound. The practices and procedures described in the following sections pertain to the National Institutes of Health and may not be universally applicable to other institutions. Administrators and/or researchers at other institutions should modify the following items as needed to reflect their individual management system and current occupational and environmental regulations.

Solutions of CP penetrate various glove materials (Laidlaw et al., 1984; Slevin et al., 1984). This factor should be taken into account when handling CP.

1. Chemical inactivation: Validated methods have been reported (Castegnaro et al., 1985).
2. Decontamination: Turn off equipment that could be affected by CP or the materials used for cleanup. If there is any uncertainty regarding the procedures to be followed or decontamination, call the NIH Fire Department (dial 116) for assistance. Consult Castegnaro et al. (1985) for details concerning decontamination of surfaces, glassware, and animal cages.
3. Disposal: It may be possible to decontaminate waste streams containing CP before disposal. For details, see Castegnaro et al. (1985). No waste streams containing CP shall be disposed of in sinks or general refuse. Surplus CP or chemical waste streams contaminated with CP shall be handled as hazardous chemical waste and disposed of in accordance with the NIH chemical waste disposal system. Nonchemical waste (e.g., animal carcasses and bedding) containing CP shall be handled and packaged for incineration in accordance with the NIH medical-pathological waste disposal system. Potentially infectious

waste (e.g., tissue cultures) containing CP shall be disinfected by heat using a standard autoclave treatment and packaged for incineration, as above. Burnable waste (e.g., absorbent bench top liners) minimally contaminated with CP shall be handled as potentially infectious waste and packaged for incineration, as above. Absorbent materials (e.g., associated with spill clean-up) grossly contaminated shall be handled in accordance with the chemical waste disposal system. Radioactive waste containing CP shall be handled in accordance with the NIH radioactive waste disposal system.

4. Storage: Store solid CP and its solutions in dark-colored, tightly closed containers, preferably under refrigeration. Avoid exposure to light and moisture. Store working quantities of CP and its solutions in an explosion-safe refrigerator in the work area.

Monitoring and Measurement Procedures Including Direct Field Measurements and Sampling for Subsequent Laboratory Analysis

1. Sampling: Biological samples are frozen (if analysis is delayed) and extracted with organic solvents from alkaline solution (Nayar et al., 1979; van den Bosch and De Voss, 1980). For large samples, such as 24 hour urine collection, concentration prior to purification on Amberlite columns is useful (Evelo et al., 1986).
2. Analysis: The current methods of choice are by gas chromatography of trifluoroacetyl (Pantarotto et al., 1974; Jardine et al., 1978; Juma et al., 1979; van den Bosch and De Voss, 1980) or heptafluorobutyl derivatives (Nayar et al., 1979), using electron capture, flame ionization, or nitrogen-phosphorus detectors. The latter is the most sensitive procedure, with detection limits of 10 ng/ml. A highly specific method (involving, however, considerable sample preparation) is isotope dilution mass spectrometry (Jarman et al., 1975; Bahr et al., 1980). Simultaneous analysis for active metabolites of CP in addition to the parent compound has been accomplished with gas (Jardine et al., 1978) and thin layer chromatography (Norpoth et al., 1975). Other less frequently used methods are HPLC with detection by ultraviolet at 200 nm (not sensitive enough for biological media; Kensler et al., 1979), infrared analysis (ditto), and a modification of the 4(4'-nitrobenzyl)pyridine colorimetric method (CP reacts with this reagent only after preliminary hydrolysis to nor-nitrogen mustard).

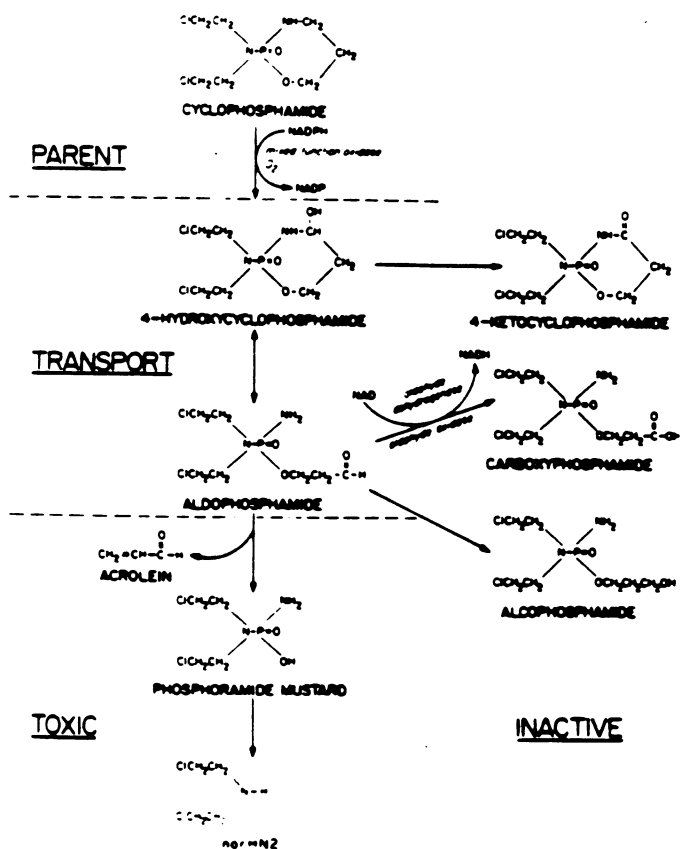
Biological Effects (Animal and Human)

1. Absorption: CP is absorbed from the intestinal tract, by parenteral injection, and through the intact skin (in organic solution) (Hirst et al., 1984; Wagner and Fenneberg, 1984). It is also transmitted transplacentally.

Distribution and pharmacokinetics: There are no data on distribution of the intact molecule. The ^{14}C label of side chain labelled CP is rapidly distributed throughout the human body after intravenous injection; 70% of the label is subsequently excreted in the urine (Bagley et al., 1973).

Pharmacokinetic data have been published (Grochow and Colvin, 1983; Wagner and Fenneberg, 1984).

Metabolism and excretion: The metabolism of CP has been worked out in detail and has been reviewed (Colvin, 1978; Domeyer and Sladek, 1980; Przybylski, 1982). It is outlined as follows:



(From Domeyer and Sladek, 1980.)

All intermediates have been identified as metabolites in animals, and most of them in man. 4-Hydroxy CP and its tautomer aldophosphamide are regarded as transport forms and may not be active in themselves, although 4-hydroxy CP is regarded as representing the major contributor to cytotoxic effects of CP by being the major cell entrant (Powers and Slader, 1983). Additional metabolites, which have not as yet been fitted into the above pathways, are 2-chloroacetaldehyde (Shaw et al., 1983), and three phosphorus-free oxazolin-2-one ring structures (Chan et al., 1986). Of these, the highly reactive 2-chloroacetaldehyde may be responsible at least partly for the toxic effects of CP in the bladder. It is noteworthy that CP

recovered from the urine of patients who were administered racemic CP is levorotatory, indicating a stereo-selective metabolism of CP (Cox et al., 1976).

The metabolites which are probably the active alkylating agents, responsible for toxic and carcinogenic effects, are phosphoramid mustard, nor-nitrogen mustard, and acrolein. Urinary excretion products are mainly unmetabolized CP, 4-hydroxy CP, acrolein (both as conjugation products with sulfhydryl compounds such as glutathione), 4-keto CP, phosphoramid mustard, and carboxy-phosphamid (Giles, 1979; Wagner et al., 1980).

Toxic effects: Acute LD50s have been reported as follows (in mg/kg): mouse, 580 iv, 400 ip, 780 oral; rat, 196 iv, 182 ip, 180 oral; guinea pig, 400 iv; dog, 40 iv, 44 oral. These figures apply to adult animals; it should be noted that in the newborn rat (the only species thus studied) toxicity is highly age-dependent: the LD50 (sc) is 35 and 320 mg/kg one and 30 days after birth (Stekar, 1973) which is in line with even higher embryotoxicity. The reason for and significance of this effect, particularly as it may apply to other species, is not known.

Toxic side effects of CP in animals and as a result of chemotherapy have been reviewed (Colvin and Hilton, 1981). There is depression of the hematopoietic system, leukopenia, alopecia, and hemorrhagic cystitis, probably due to renal excretion of 4-hydroxy metabolites and of acrolein (Brock et al., 1981). In early studies the urinary tract damage due to hemorrhagic cystitis was dose limiting, but it can be prevented by simultaneous administration of Mesna (sodium-2-mercaptoethane sulfonate) (Grochow and Colvin, 1983).

Early toxic signs in man are nausea, vomiting, and water retention. Compared with other alkylating agents, CP has exceptionally high immunosuppressive activity which makes it the agent of choice in the treatment of autoimmune diseases and in renal and bone marrow transplants. The biochemical lesion produced by CP metabolites is inhibition of thymidine incorporation into tissue DNA and of RNA synthesis.

Carcinogenic effects: These have been reviewed in detail (IARC, 1981) and it is concluded that there is sufficient evidence for carcinogenicity in mice, rats, and man. Examples are: In a life time experiment, administration of 0.63, 1.25, or 2.5 mg/kg of CP 5 times per week in drinking water resulted in dose-dependent incidence of carcinoma of the bladder and tumors of the lymphoid and the hematopoietic system (Schmahl and Habs, 1979). Pulmonary tumors were induced in mice after intraperitoneal administration, but only in doses 300 times higher than those of nitrogen mustard producing an equivalent response (Shimkin et al., 1966). Application of CP (0.5 ml of a 0.4%

acetone solution) to the skin of mice resulted in a significant incidence of mammary tumors (Matsuyama et al., 1966). There are several reports of tumor induction in patients treated with CP for nephrosis or malignancies; most often seen were bladder carcinomas. These reports are difficult to evaluate since in most cases there was concomitant radiation or other chemotherapy.

6. Mutagenic and teratogenic effects: CP is mutagenic after activation in the Ames test and against E. coli and Drosophila (Ellenberger and Mohn, 1975; Seino et al., 1978), and teratogenic and embryotoxic in the chick, rabbit, mouse, and rat. Since metabolites of CP (4-hydroxy CP, phosphoramid mustard) are mutagenic without activation being required, these are presumably the direct mutagens and teratogens. Acrolein may also play a role in the mutagenicity and teratogenicity of CP (Hales, 1982).

Emergency Treatment

1. Skin and eye exposure: For skin exposure, remove contaminated clothing and wash skin with soap and water. Skin should not be rinsed with organic solvents or scanned with UV light. Since CP is readily absorbed through the skin, avoid rubbing of skin or increasing its temperature. For eye exposure, irrigate immediately with copious quantities of running water for at least 15 minutes. Obtain ophthalmological evaluation.
2. Ingestion: Drink plenty of water or milk. Induce vomiting. Refer for gastric lavage.
3. Inhalation: Remove victim promptly to clean air. Administer rescue breathing if necessary.
4. Refer to physician. Consider treatment for pulmonary irritation.

References

- Althouse, R., J. Huff, L. Tomatis, and J. Wilbourn. 1980. An evaluation of chemicals and industrial processes associated with cancer in humans based on human and animal data. IARC monograph volumes 1 to 20. Cancer Res 40:1-12.
- Arnold, H., and H. Klose. 1960. Die Hydrolyse hexacyclischer N-Loth phosphamid ester im gepufferten System. [The hydrolysis of hexacyclic phosphamide mustard esters in a buffered system.] Arzneimittel-Forsch 10:288-291.
- Bagley, C.M. Jr., F.W. Bostick, and V.T. DeVita Jr. 1973. Clinical pharmacology of cyclophosphamide. Cancer Res 33:226-233.
- Bahr, U., H.-R. Schulten, O.R. Hommes, and F. Aerts. 1980. Determination of cyclophosphamide in urine, serum and cerebrospinal fluid of multiple sclerosis patients by field desorption mass spectrometry. Clin Chim Acta 103:183-192.

- Benvenuto, J.A., R.A. Anderson, K. Kerkof, R.G. Smith, and T.L. Loo. 1981. Stability and compatibility of antitumor agents in glass and plastic containers. *Am J Hosp Pharm* 38:1914-1918.
- Brock, N., J. Pohl, and J. Stekar. 1981. Studies on the urotoxicity of oxazaphosphorine cytostatics and its prevention. I. Experimental study on the urotoxicity of alkylating compounds. *Europ J Cancer* 17:595-607.
- Brooke, D., R.J. Bequette, and E. Davis. 1973. Chemical stability of cyclophosphamide in parenteral solutions. *Am J Hosp Pharm* 30:134-137.
- Castegnaro, M., J. Adams, M.A. Armour, J. Barek, J. Benvenuto, C. Confalonieri, U. Goff, S. Ludeman, D. Reed, E.B. Sansone, and G. Telling. 1985. Laboratory Decontamination and Destruction of Carcinogens in Laboratory Wastes: Some Antineoplastic Agents. IARC Scientific Publications No. 73. World Health Organization, Geneva, Switzerland.
- Chan, K.K., S.-C. Hong, E. Watson, and S.-K. Deng. 1986. Identification of new metabolites of phosphoramidate and non-nitrogen mustards and cyclophosphoramidate in rat urine using ion cluster techniques. *Biomed Environ Mass Spectrom* 13:145-154.
- Colvin, M. 1978. A review of the pharmacology and clinical use of cyclophosphamide in: Pinedo, M.M. (ed), *Clinical Pharmacology of Anti-Neoplastic Drugs*. *Appl Methods in Oncol* 1:245-261.
- Colvin, M., and J. Hilton. 1981. Pharmacology of cyclophosphamide and metabolites. *Cancer Treat Rep* 65(Suppl 3):89-95.
- Cox, P.J., P.B. Farmer, M. Jarman, M. Jones, W.J. Stec, and R. Kinas. 1976. Observations on the differential metabolism and biological activity of the optical isomers of cyclophosphamide. *Biochem Pharmacol* 25:993-996.
- Domeyer, B.E., and N.E. Sladek. 1980. Kinetics of cyclophosphamide biotransformation in vivo. *Cancer Res* 40:174-180.
- Ellenberger, J., and G. Mohn. 1975. Mutagenic activity of cyclophosphamide, ifosfamide and trofosfamide in different genes of Escherichia coli and Salmonella typhimurium after biotransformation through extracts of rodent liver. *Arch Toxicol* 33:225-240.
- Evelo, C.T.A., R.P. Bos, J.G.P. Peters, and P.T. Henderson. 1986. Urinary cyclophosphamide assay as a method for biological monitoring of occupational exposure to cyclophosphamide. *Int Arch Occup Environ Health* 58:151-155.
- Gallelli, J.R. 1967. Stability studies of drugs used in intravenous solutions. Part 1. *Am J Hosp Pharm* 24:425-433.
- Giles, P.M. 1979. The biosynthesis of 3-hydroxypropylmercapturic acid from cyclophosphamide. *Xenobiotica* 9:745-762.
- Grochow, L.B., and M. Colvin. 1983. Clinical pharmacokinetics of cyclophosphamide in: Ames, M.M., G. Powis, and J.S. Kovach (eds), *Pharmacokinetics of Anticancer Agents in Humans*, Ch. 6, Elsevier, New York.

- Hales, B.F. 1982. Comparison of the mutagenicity and teratogenicity of cyclophosphoramide and its active metabolites, 4-hydroxycyclophosphoramide, phosphoramide mustard, and acrolein. *Cancer Res* 42:3016-3021.
- Hirst, M., D.G., Mills, S. Tse, and L. Levin. 1984. Occupational exposure to cyclophosphoramide. *Lancet* (1) 186-188.
- IARC. 1981. International Agency for Research on Cancer. Cyclophosphamide. IARC Monographs 26:165-202.
- Jardine, I., C. Fenselau, M. Appler, M.-N. Kan, R.B. Bundrett, and M. Colvin. 1978. Quantitation by gas-chromatography-chemical ionization mass spectrometry of cyclophosphoramide, phosphoramide mustard, and nor-nitrogen mustard in the plasma and urine of patients receiving cyclophosphoramide therapy. *Cancer Res* 38:408-415.
- Jarman, M., E.D. Gilby, A.B. Foster, and P.K. Bondy. 1975. The quantitation of cyclophosphamide in human blood and urine by mass spectrometry--stable isotope dilution. *Clin Chim Acta* 58:61-69.
- Juma, F.D., H.J. Rogers, and J.R. Trounce. 1979. Pharmacokinetics of cyclophosphamide and alkylating activity in man after intravenous and oral administration. *Brit J Clin Pharmacol* 8:209-217.
- Kensler, T.T., R.J. Behme, and D. Brooke. 1979. High performance liquid chromatographic analysis of cyclophosphamide. *J Pharm Sci* 68:172-174.
- Laidlaw, J.L., T.H. Connor, J.L. Theiss, R.W. Anderson, and T.S. Matney. 1984. Permeability of latex and polyvinyl chloride gloves to 20 antineoplastic drugs. *Am J Hosp Pharm* 41:2618-2623.
- Laine, E., V. Tuominen, H. Jalonen, and P. Kahela. 1983. Effect of storage conditions on structure of cyclophosphamide. *Acta Pharm Fenn* 92:243-248; *Chem Abstr* 101:28154f.
- Matsuyama, M., A. Maekawa, and T. Nakamura. 1966. Biological studies on anticancer agents. II. Effect of percutaneous application. *Gann* 57:295-298.
- Mruzek, M.N., and I.C. Shaw. 1984. A mass spectral study of cyclophosphamide with special reference to differences in isomeric structures. *Biomed Mass Spectrom* 11:360-366.
- Nayar, M.S.B., L.-Y. Lin, S.H. Wan, and K.K. Chan. 1979. Gas chromatographic analysis of cyclophosphamide in plasma and tissues using nitrogen-phosphorus detection. *Anal Ltrs* 12:905-915.
- Norpoth, K., H.W. Addicks, U. Witting, G. Müller, and H. Raidt. 1975. Quantitative Bestimmung von Cyclophosphamid, Ifosfamid und Trofosfamid sowie ihrer stabilen Metabolite auf der DC-Platte mit 4-Pyridinaldehyd-2-benzothiazolyl-hydrazon (PBH). [Quantitative determination of cyclophosphamide, ifosfamide and trofosfamide and their stable metabolites by TLC with PBH]. *Arzneimittel-Forsch* 25:1331-1336.
- Pantarotto, C., A. Bossi, G. Belvedere, A. Martini, M.G. Donelli, and A. Frigerio. 1974. Quantitative GLC determination of cyclophosphamide and isophosphamide in biological specimens. *J Pharm Sci* 63:1554-1558.

- Powers, J.F., and N.E. Sladek. 1983. Cytotoxic activity relative to 4-hydroxycyclophosphamide and phosphoramidate mustard concentrations in the plasma of cyclophosphamide-treated rats. *Cancer Res* 43:1101-1106.
- Przybylski, M. 1982. Identification of metabolism pathways of anticancer drugs by high-pressure liquid chromatography in combination with field desorption mass spectrometry. *Arzneimitt-Forsch* 32:995-1012.
- Sato, T., H. Ueda, K. Nakagawa, and N. Bodor. 1983. Asymmetric synthesis of enantiomeric cyclophosphamide. *J Org Chem* 48:98-101.
- Schmähl, D., and M. Habs. 1979. Carcinogenic action of low-dose cyclophosphamide given orally to Sprague-Dawley rats in a life-time experiment. *Int J Cancer* 23:706-712.
- Schulten, H-R. 1974. Field desorption mass spectrometry of metabolites of the anticancer drug cyclophosphamide. *Biomed Mass Spectrom* 1:223-230.
- Seino, Y., M. Nagao, T. Yahagi, A. Hoshi, T. Kawachi, and T. Sugimura. 1978. Mutagenicity of several classes of antitumor agents to *Salmonella typhimurium* TA98, TA100, and TA92. *Cancer Res* 38:2148-2156.
- Shaw, I.C., M.I. Graham, and A.E.M. McLean. 1983. 2-chloroacetaldehyde, a metabolite of cyclophosphamide in the rat. *Xenobiotica* 13:433-437.
- Shimkin, M.B., J.H. Weisburger, E.K. Weisburger, N. Gubareff, and V. Sontzeff. 1966. Bioassay of 29 alkylating chemicals by the pulmonary-tumor response in Strain A mice. *J Nat Cancer Inst* 36:915-935.
- Slevin, M.L., L.M. Ang, A. Johnston, and P. Turner. 1984. The efficiency of protective gloves used in the handling of cytotoxic drugs. *Cancer Chemother Pharmacol* 12:151-153.
- Steckar, J. 1973. Teratogenicity of cyclophosphamides in newborn rats. *Arzneimitt-Forsch* 23:922-923.
- van den Bosch, N., and D. DeVoss. 1980. Some aspects of the gas-liquid chromatographic analysis of cyclophosphamide in plasma. *J Chromatog* 183:49-56.
- Wagner, T., D. Heydrich, G. Voelcker, and H.J. Hohorst. 1980. Über Blutspiegel und Urin-Ausscheidung von aktiviertem Cyclophosphamid und seinen Deaktivierungsprodukten beim Menschen. [Blood levels and urinary excretion of activated cyclophosphamide and its products of deactivation in man.] *J Cancer Res Clin Oncol* 96:79-92.
- Wagner, T., and K. Fenneberg. 1984. Pharmacokinetics and bioavailability of cyclophosphamide from oral formulations. *Arzneimitt-Forsch* 34:313-316.
- Wantland, L.R., and S.D. Hersh. 1979. High performance liquid chromatographic assay of cyclophosphamide in raw material and parenteral dosage forms. *J Pharm Sci* 68:1144-1146.
- Zon, G., S.M. Ludeman, and W. Egan. 1977. High-resolution nuclear magnetic resonance investigations of the chemical stability of cyclophosphamide and related phosphoramidate compounds. *J Am Chem Soc* 99:5785-5795.